Oxidation of Polycyclic Aromatic Hydrocarbons Catalyzed by Iron Tetrasulfophthalocyanine FePcS: Inverse Isotope Effects and Oxygen Labeling Studies

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Iron(III) tetrasulfophthalocyanine (FePcS) was shown to catalyze the oxidation of polycyclic aromatic hydrocarbons by H_2O_2 . Benzo[a]pyrene and anthracene were converted to the corresponding quinones while biphenyl-2,2'-dicarboxylic acid was the main product of phenanthrene oxidation. The mechanism of the anthracene oxidation by H2O2 in the presence of FePcS or by KHSO₅ with iron(III) mesotetrakis(3,5-disulfonatomesityl)porphyrin (FeTMPS) (see Figure 1 for catalyst structures) has been investigated in details by using kinetic isotope effects (KIEs) and ¹⁸O labeling studies. KIEs measured on the substrate consumption in the competitive oxidation of $[H_{10}]$ anthracene and [D₁₀]anthracene by FePcS/H₂O₂ and FeTMPS/KHSO₅ were essentially the same, 0.75 ± 0.02 and 0.76 ± 0.06 , respectively. These inverse KIEs on the first oxidation step can be explained by the sp²-to-sp³ hybridization change during the addition of an electrophilic oxoiron complex to the sp² carbon center of anthracene to form a σ adduct (this inverse KIE being enhanced by stronger stacking interactions between the perdeuterated substrate with the macrocyclic catalyst). Although the first oxidation step seems to be the same, different distribution of the oxidation products of anthracene and very different $^{18}\mbox{O}$ incorporation into anthrone and anthraquinone in catalytic oxidations performed in the presence of H₂¹⁸O suggested that different active species should be responsible for anthracene oxidation in both catalytic systems. All the results obtained are compatible with an involvement of TMPSFeV=O (or TMPS+Fe^{IV}=O), having two redox equivalents above the iron(III) state of the metalloporphyrin precursor, while PcSFe^{IV}=O (one redox equivalent above Fe^{III} state of FePcS) was proposed to be the active species in the metallophthalocyanine-based system.

Introduction

Polycyclic aromatic hydrocarbons (PAH) belong to environmental pollutants because of their rather slow biodegradation. PAHs can be metabolized by bacteria, fungi or algae. Three main degradation pathways have been identified for the microbial catabolism of polycyclic aromatic hydrocarbons^[1]. Cytochrome P-450 monooxygenases catalyze the formation of arene oxides, which are then transformed to phenol or epoxy-diol derivatives. Ligninases are capable of oxidizing PAH to quinones followed by ring cleavage^[2]. For example, benzo[a]pyrene was oxidized to a mixture of 1,6-, 3,6- and 6,12-quinones^[3], pyrene to 1,6- and 1,8-quinones^[4] and anthracene to 9,10-anthraquinone^[2]. The third enzymatic degradation process involves dioxygenase enzymes to incorporate both atoms of molecular oxygen into the aromatic ring to form cis-dihydrodiols. After a dehydrogenase-catalyzed rearomatization to produce catechols, a subsequent enzymatic cleavage by intradiol or extradiol dioxygenases gives cis, cis-muconic acid or 2-hydroxymuconic semialdehyde as products of aromatic cycle cleavage, respectively^[5]. Although biodegradation of PAHs is the major decontamination process, along with photochemical oxidation with dioxygen^[6], microorganisms capable of rapidly and efficiently mineralizing recalcitrant and potentially carcinogenic PAHs needs to be found[1]. Chemical oxidation of PAH^[7] needs elevated temperatures and/or powerful oxidants, e.g. chromium compounds which constitute environmental problems. For example, phenanthrene has been oxidized to phenanthrenequinone by CrO₃ at 55°C (yield 80%)^[8], by the system Ce^{IV}/H₂SO₄/(NH₄)₂SO₄ at 50°C (yield 42%)^[9], by KMnO₄/Al₂O₃/H₅IO₆ (yield 25%)^[10]. Arenes can be oxidized to quinones by H₂O₂/hexafluoroacetone at 70°C^[11]. In this latter case, biphenyl-2,2'-dicarboxylic acid was the only isolated oxidation product of phenanthrene^[11]. Obviously, there is a need for environmental-friendly chemical catalysts to oxidize PAH. An alternative approach using biomimetic metalloporphyrin catalysts^[12] has been shown to be useful in the oxidation of 2-methylnaphthalene^[12a]. Phenanthrene and naphthalene were oxidized by a porphyrinruthenium/2,6-dichloropyridine N-oxide system to give the corresponding quinones with 40% and 29% yields, respectively^[13]. Recently, the oxidation of naphthalene and methylnaphthalenes by peracetic acid catalyzed by octanitrophthalocyanine Mn^{III} and Fe^{II} complexes has been reported^[14].

We recently developed a new efficient catalytic system for the oxidative degradation of polychlorinated phenols based on the association of iron(III) or manganese(III) complexes of tetrasulfophthalocyanine (FePcS, MnPcS) and the rather inexpensive and "green" oxidant H₂O₂[15][16]. This biomimetic system is able to cleave the aromatic cycle of 2,4,6trichlorophenol with the formation of chloromaleic acid as the main product^[15]. Besides FePcS and MnPcS, CoPcS is also highly efficient in the oxidation of catechols^[17]. Having on hands these efficient catalysts for the oxidation of recalcitrant pollutants such as chlorinated phenols, we decided to investigate the activity of these catalysts in the oxidation of other poorly biodegradable molecules. Here we report the catalytic oxidation of polycondensed aromatics (mainly anthracene, but also phenanthrene and benzo[a]pyrene) by hydrogen peroxide or potassium monopersulfate (KHSO₅) and FePcS as catalyst [FeTMPS, a water-soluble iron(III) porphyrin, was also used for comparison]. Detailed mechanistic studies involving kinetic isotope effects (KIEs) and ¹⁸O labeling experiments are also reported.

Figure 1. Structures of iron(III) complexes of tetrasulfophthalocy-anine (FePcS) and *meso*-tetrakis(3,5-disulfonatomesityl)porphyrin

Iron tetrasulfophthalocyanine (FePcS)

Iron meso-tetrakis(3,5-disulfonatomesityl)porphyrin (FeTMPS)

Results

Products and Reactivity Order in PAH Oxidations by $FePcS/H_2O_2$

We checked the oxidations of benzo[a]pyrene, phenanthrene and anthracene, performed in a MeCN/H2O mixture (3:1, v/v) using KHSO₅ and H₂O₂ as oxidants. The final concentrations of oxidant and substrate were 5 mm and 1 mm, respectively. The catalyst concentrations were 10 μ M, 37 μ M and 100 μ M corresponding to 1, 3.7 and 10% catalyst/substrate ratios, respectively. The data of Table 1 indicated that FePcS was also an efficient catalyst for the oxidation of these polyaromatic hydrocarbons. The oxidation of benzo[a]pyrene by FePcS/KHSO₅ (run 1) and by FePcS/H₂O₂ (run 4) at pH = 7 was complete within in a few minutes leading to an orange material isolated by precipitation with water. DCI-CH₄ mass spectrometry analysis gave peaks at 282 (M⁺, 100), 254 [(M - CO)⁺, 17.4], 226 $[(M - 2 CO)^+, 17.4]$ indicating the final product was a quinone (or a mixture of quinone isomers) with a molecular weight equal to 282. Three components were isolated by dry column chromatography (neutral Al₂O₃, benzene) after oxidation of benzo[a]pyrene in conditions used for run 4 of Table 1 and characterized by UV spectroscopy according to ref.^[18] (Scheme 1). The UV/Vis spectra for all three quinones were in an excellent agreement with the data previously published^[18]. Based on reported extinction coefficients, the distribution of quinones was as follows: 6,12benzo[a]pyrenedione (32%, first eluted product from column), 1,6- benzo[a]pyrenedione (44%, second eluted product) and 3,6- benzo[a]pyrenedione (24%, third eluted product). The total yield for all three quinones was quantitative

Table 1. Oxidation of polyaromatic hydrocarbons PAH by H_2O_2 and $KHSO_5$ catalyzed by sulfonated phthalocyanineiron(III) $FePcS^{[a]}$

Run	Oxidant	pН	% Cat./Sub.	Conver	sion [%]	
				.1 min	5 min	60 min
Benzo[a]pyrene oxidation						
1	$KHSO_5$	2	1	50	97	100
2	$KHSO_5$	7	1	14	41	70
2 3	H_2O_2	7	1	11	20	20
4	H_2O_2	7	3.7	43	100	100
	2 2	Anthracene oxidation				
5	H_2O_2	4.4	10	12	40	96
6	H_2O_2	7	10	100		, 0
7	H_2O_2	7	3.7	57	74	86
8	H_2O_2	7	1	15	28	32
-	2 - 2	Anthracene oxidation by FeTMPS/KHSO ₅				
9	KHSO ₅	7	3.7	11	33	100
		Phenanthrene oxidation				
10	KHSO ₅	7	3.7	28	40	64
11	H_2O_2	5.3	3.7	10	24	35
12	H_2O_2	5.3	10	n.d.	n.d.	66
13	H_2O_2	5.5	10	32	46	64
14	H_2O_2	5.9	10	n.d.	45	60
15	H_2O_2	4.4	10	n.d.	24	80

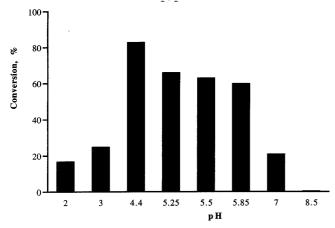
[[]a] n.d.: not determined.

The H_2O_2 oxidation of phenanthrene is pH-dependent. Using a 10% catalyst/substrate ratio the highest conversion of phenanthrene was obtained at pH = 4.4 (conversion be-

Scheme 1. Oxidation products of benzo[a]pyrene by FePcS/H₂O₂

ing rather low at pH = 7) (Figure 2). But it should be noted that anthracene was quickly and completely oxidized within 1 min at pH = 7 (Table 1, run 6) while 1 hour was necessary to reach 96% conversion at pH = 4.4 (the pH optimum for phenanthrene). At pH = 7 the order of substrate reactivity was in agreement with oxidation potentials: benzo[a]pyrene > anthracene > phenanthrene (the ionization potentials being 7.2, 7.4 and 8.1 eV, respectively^[2]).

Figure 2. pH dependence of phenanthrene oxidation by FePcS/



 $^{\rm [a]}$ Reaction conditions: [catalyst] = 0.1 mm, [oxidant] = 5 mm, [substrate] = 1 mm, acetonitrile/50 mm phosphate buffer at pH = 7 (3:1, v/v). Reaction time 60 min.

The FePcS-catalyzed oxidation of anthracene by $\rm H_2O_2$ resulted in the formation of anthrone (tautomeric form of 9-hydroxyanthracene), oxanthrone (tautomeric form of anthrahydroquinone) and anthraquinone (Scheme 2). The yields of oxanthrone (5%) and anthraquinone (25%) were determined after 1 h by HPLC using calibration curve and molar extintion coefficients obtained with authentic samples.

Biphenyl-2,2'-dicarboxylic acid (product of cleavage of one aromatic cycle) was the main oxidation product of phenanthrene, 9,10-phenanthrenequinone being the second main product. Three other minor products of the oxidative degradation of phenanthrene were also identified by GC-MS analysis: 2-formyl-2'-hydroxybiphenyl, 2-carboxy-2'-hydroxybiphenyl and 2,2'-biphenylcarbolactone (Scheme 3). So, the FePcS/H₂O₂ system is able to cleave the aromatic cycle of phenanthrene and to perform oxidative decarboxylation.

Oxidation of Anthracene by FeTMPS/KHSO₅

We also checked the oxidation of anthracene by FeTMPS/KHSO₅ performed in an MeCN/25 mm phosphate buffer mixture (3:1, v/v) at pH = 7. The concentrations of oxidant, substrate and catalyst were 5 mm, 1 mm and 37 µM, respectively. The anthracene conversion was complete after 60 min (run 9, Table 1). The yields of oxanthrone and anthraquinone were determined by HPLC. Differing from the FePcS/H₂O₂ mediated oxidation, the main product with FeTMPS/KHSO₅ was oxanthrone (yield by HPLC = 60%), while only 3% of anthraguinone were found. Isolation of reaction products from a large scale anthracene oxidation by FeTMPS/KHSO5 followed by NMR analysis gave 43% yield for oxanthrone, 16% yield for anthraquinone and 14% yield of non-identified compounds (probably coupling products). A partial oxidation of oxanthrone to anthraquinone might have occured during the sample preparation for NMR analysis, explaining the discrepancy with the HPLC analysis of the reaction mixture. This rather unexpected result, oxanthrone, an easily oxidizable compound, as main reaction product in the presence of strong oxidants, stimulated us for a detailed mechanistic study of anthracene oxidation by both catalytic systems, FePcS/H₂O₂ and FeTMPS/KHSO₅.

Scheme 2. Oxidation products of anthracene by FePcS/H₂O₂ or FeTMPS/KHSO₅

Scheme 3. Oxidation products of phenanthrene by FePcS/H₂O₂

Stability of Anthrone, Oxanthrone and Anthraquinone in the Presence of FePcS/H₂O₂ and FeTMPS/KHSO₅

In these experiments we used the same reaction conditions as those for anthracene oxidation: a MeCN/50 mm phosphate buffer pH = 7 reaction medium (3:1, v/v), [catalyst] = 37 μ M, [oxidant] = 5 mM, [substrate] = 1 mM, except for anthraquinone (its concentration was reduced to 0.5 mm because of its low solubility in the reaction mixture). Under these reaction conditions the conversions of anthrone in the presence of FePcS/H₂O₂ and FeTMPS/KHSO₅ were 23% and 38%, respectively, after 1 h, indicating that anthrone oxidation was slightly faster with FePcS/KHSO₅. Oxanthrone was easily oxidized to anthraquinone by FePcS/ H₂O₂: the conversion was 90% after only 5 min. Again, oxanthrone was relatively stable with FeTMPS/KHSO₅: the conversions were only 18 and 23% after 30 and 60 min, respectively. Finally, we checked that anthraquinone was stable in both catalytic systems. There was no observable conversion after 60 min with FeTMPS/KHSO₅ and only 10% with FePcS/H₂O₂. These results on stability of products of anthracene oxidation were in good agreement with the composition of the reaction products for both catalytic systems.

Method of Determination of Kinetic Isotope Effects

Kinetic isotope effects (KIEs) can be determined in intraor intermolecular competitive experiments by the analysis of the ratios of reaction products resulting from C-H versus C-D bond transformations. Such analyses are reliable when the oxidation products, different only in their isotopic substitution, are primary oxidation products and are stable in the reaction conditions. Since this is not a case in PAH oxidations by FePcS/H₂O₂ system we decided to determine the KIE values by analyzing of the competitive consumption of the deuterated versus the non-deuterated substrate. In this case intermolecular KIE values of the initial oxidation reaction can be obtained. In order to determine these KIEs concentrations of labeled and unlabeled substrates should be determined before and after the oxidation reaction by monitoring the isotopic composition of the initial reaction mixture, the substrate conversion and the isotopic composition of the remaining substrate after reaction. Isotopic compositions of substrates were determined by

GC-MS analyses and substrate conversions were obtained by HPLC analyses (see Experimental Section). Equations to calculate KIEs depend on a reaction order in substrate [19]. When the reaction is first order in substrate KIEs can be determined from equation (1) where $a_{\rm H}{}^{\rm o}$, $a_{\rm D}{}^{\rm o}$ and $a_{\rm H}$, $a_{\rm D}$ denote initial and current substrate concentrations of H-labeled and D-labeled substrates, respectively.

$$k_{\rm H}/k_{\rm D} = \log (a_{\rm H}/a_{\rm H}^{\rm o}) / \log (a_{\rm D}/a_{\rm D}^{\rm o})$$
 (1)

Determination of the Reaction Order for the Substrate in Anthracene Oxidation by FePcS/H₂O₂

The reaction order in anthracene was determined with concentrations of H_2O_2 , anthracene and FePcS being 5 mm, 1 mm and 37 μ m, respectively. The reactions were carried out in an MeCN/50 mm phosphate buffer mixture at pH = 7 (3:1, v/v). Under these conditions the catalytic reaction follows a first-order kinetic in substrate at least for 2 times the reaction half-time and KIEs could be then determined according to equation (1).

KIEs in Anthracene and Phenanthrene Oxidation by FePcS/ H_2O_2 and FeTMPS/KHSO $_5$

Analysis of the reaction mixture after 3–5 min of reaction (conversions being 40–60%) indicated a preferential consumption of [D₁₀]anthracene corresponding to an inverse isotope effect. The kinetic isotope effect in anthracene oxidation by the FePcS/H₂O₂ system was determined to be equal to 0.75 \pm 0.02. Oxidation of anthracene by FeTMPS/KHSO₅ presented the same isotope effect of 0.76 \pm 0.06 (Table 2). In contrast, oxidation of phenanthrene by both catalytic systems proceeds with no significant isotope effect: 0.96 \pm 0.04 and 1.02 \pm 0.01 for FePcS/H₂O₂ and FeTMPS/KHSO₅, respectively (Table 2).

In order to find the origin of this inverse isotope effects we prepared 9,10-dideuterioanthracene and determined the KIE value in the competitive oxidation of the mixture of 9,10-dideuterioanthracene and [H $_{10}$]anthracene. In this latter case the KIE value were 0.90 \pm 0.05 and 0.95 \pm 0.01 with the FePcS/H $_2$ O $_2$ and FeTMPS/KHSO $_5$ systems, respectively.

Table 2. Kinetic isotope effect [a] in anthracene and phenanthrene oxidation by FePcS/H $_2$ O $_2$ and FeTMPS/KHSO $_5$ at 20 °C

	KIE in the competi	KIE in the competitive oxidation of				
Catalytic system	$[H_{10}]$ anthracene/ $[D_{10}]$ anthracene ^[b] $([H_{10}]$ anthracene/9,10- $[D_2]$ anthracene)	$[H_{10}] phen anthrene/\\ [D_{10}] phen anthrene^{[c]}$				
FePcS/H ₂ O ₂ FeTMPS/ KHSO ₅	$0.75 \pm 0.02 (0.90 \pm 0.05)$ $0.76 \pm 0.06 (0.95 \pm 0.01)$	0.96 ± 0.04 1.02 ± 0.01				

^[a] These data are the mean values obtained from 3 independent oxidation reactions. - ^[b] Reaction conditions: [catalyst] = 37 μ M, [oxidant] = 5 mM, [substrate] = 1 mM, MeCN/50 mM phosphate buffer pH = 7 (3:1, v/v). - ^[c] Reaction conditions: [catalyst] = 100 μ M, [oxidant] = 5 mM, [substrate] = 1 mM, MeCN/33.3 mM phosphate buffer pH = 4.6 (1:1, v/v).

Analysis of Isotopic Composition of Products of Anthracene Oxidation

Anthrone, oxanthrone and anthraquinone were identified as oxidation products of anthracene by NMR and GC-MS techniques. GC-MS analysis of the reaction mixture indicated that anthrone, the primary oxidation product, was enriched in the deuterated derivative as expected from the inverse isotope effect on the first step of anthracene oxidation. In contrast, anthraquinone, a second oxidation product, was enriched in unlabeled product indicating a KIE value > 1 during the oxidation of anthrone to this quinone. Oxanthrone was unstable under GC-MS analysis conditions and its isotopic composition could not be determined. The molecular peak of deuterated anthrone formed after 3 min of reaction was 202 corresponding to a [D₈]containing molecule, not a [D₉]-molecule as expected for 9hydroxyanthracene. This result suggested that the equilibrium between the two tautomeric forms, anthrone and 9hydroxyanthracene, was rather fast (Scheme 4). Consequently, a putative σ adduct with a Fe-O-arene bond, which should stabilize the 9-hydroxyanthracene tautomer, could not be a long-lived intermediate as previously observed in the metalloporphyrin-catalyzed oxidation of 2methylnapthalene[12a].

¹⁸O-Labeling Experiments

Oxidations of organic substrates catalyzed by metalloporphyrin^{[12][20][21]} and metallophthalocyanine^[16] complexes in the presence of ¹⁸O-labeled water or dioxygen provided valueable mechanistic information. In order to investigate the mechanism of these catalytic PAH oxidations we performed the oxidation of anthracene by FePcS/H₂O₂ and FeTMPS/KHSO₅ in reaction mixtures containing ¹⁸O-lab-

eled water. In control experiments we checked that light oxygen atoms of anthraquinone did not exchange with heavy oxygen atoms of ¹⁸O-labeled water under the used reaction conditions, i.e. a mixture of acetonitrile/50mm phosphate buffer (3:1, v/v) at pH = 7. Also, KHSO₅ itself does not exchange its oxygen atoms with water^[20c]. FePcSmediated oxidations of anthracene were performed under standard conditions (run 7 of Table 1) either under air or under nitrogen, using a 50:50 mixture of [H₁₀]anthracene and [D₁₀]anthracene. The oxygen atom distribution was analyzed in both deuterated and non-deuterated products for each experiment. In all cases, the oxygen atom distribution for a defined product was exactly the same. For example, the ¹⁸O incorporation into anthraquinone was followed by determination of the molecular ion percentages at 208/216 [M], 210/218 [M + 2] and 212/220 [M + 4] corresponding to the incorporation of none, one or two ¹⁸O oxygen atoms in [H₈]anthraquinone and [D₈]anthraquinone, respectively. In both cases, under air or under nitrogen, anthrone, the primary oxidation product, contained only a light oxygen atom. This result indicates that this oxygen atom originated from H₂O₂, the non-labeled oxidant. In contrast, both ¹⁶O and ¹⁸O atoms were incorporated into anthraquinone. When the reaction was performed under air, the molecular cluster consisted of M (68%) and M + 2 (32%) peaks, meaning that the ¹⁸O atom was unambiguously only located in one carbonyl group of this quinone. If ¹⁸O would occupied both carbonyl positions, the molecular cluster would have consisted on three peaks: M, M + 2 and M + 4. So, anthraquinone contained 100% of ¹⁶O in one carbonyl position, 36% of ¹⁶O and 64% of ¹⁸O in another one (Figure 3). When the reaction was performed under nitrogen, again, only two peaks were found in the molecular cluster: M (56%) and M + 2 (44%). Consequently, 16 O incorporation in one carbonyl position was complete, another carbonyl position was enriched with heavy oxygen: 12% of ¹⁶O and 88% of ¹⁸O (Figure 3). This more significant incorporation of ¹⁸O at the second carbonyl position produced in the absence of dioxygen suggests a possibility of O2 involvement into oxidation pathway from anthrone to anthraquinone.

We also studied the ¹⁸O incorporation from labeled water into the oxidation products of anthracene mediated by FeTMPS/KHSO₅. In this latter case, the mass spectrum of anthrone indicated the incorporation of ¹⁸O atom, unlike with the FePcS/H₂O₂ oxidation. The percentage of ¹⁸O found in anthrone was 63%, ¹⁶O incorporation being only 37% (Figure 3). The MS molecular cluster of anthraquinone consisted of 3 peaks: M (16%), M + 2 (59%) and M

Scheme 4. Deuterium exchange during the formation of anthrone in aqueous medium

Figure 3. 18 O incorporation into anthrone and anthraquinone during the oxidation of anthracene by FePcS/H₂O₂ and FeTMPS/KHSO₅ in the presence of H₂ 18 O

+ 4 (25%), indicating that ¹⁸O was incorporated in both carbonyl groups of the quinone (the amount of ¹⁶O and ¹⁸O being 45 and 55%, respectively) (Figure 3). From these labeled water experiments we concluded that active species involved in the oxidation of anthracene by FePcS/H₂O₂ or by FeTMPS/KHSO₅ must be different.

Discussion

We found that FePcS associated with H₂O₂ can be useful in oxidation of polycyclic aromatic compounds. Anthracene was used as substrate to perform kinetic isotope studies and oxygen labeling experiments. Then we compared the results of the catalytic oxidations of anthracene by FePcS/H₂O₂ with those obtained with FeTMPS/KHSO₅ in order to gain a more complete view on the mechanisms of these oxidation reactions catalyzed by two related iron macrocyclic complexes. High-valent oxometal complexes have been proposed as active intermediates in hydrocarbon oxygenations (hydroxylations and epoxidations) using KHSO₅, NaOCl or mCPBA as oxidants and porphyriniron or -manganese compounds as catalysts [20][21][22]. We recently proposed a nucleophilic peroxoiron(III) complex of tetrasulfophthalocyanine, PcSFe^{III}OOH, as active species responsible for the oxidative cleavage of quinones generated during the oxidation of 2,4,6-trichlorophenol by FePcS/H₂O₂^{[15][16]}.

In the present work, several important differences must be noted in anthracene oxidations performed by FePcS/ H₂O₂ or FeTMPS/KHSO₅. These differences are (i) a different composition of oxidation products depending on the oxidation system used (Scheme 2), (ii) a different reactivity of oxanthrone, which is relatively stable in the presence of FeTMPS/KHSO₅, but very quickly oxidized by FePcS/ H₂O₂, (iii) and very different ¹⁸O incorporations into anthrone and anthraquinone in experiments performed in the presence of H₂¹⁸O (Figure 3). On the other side, both catalytic systems, FePcS/H₂O₂ and FeTMPS/KHSO₅, showed essentially the same KIE values in the oxidation of

anthracene (KIE = 0.75 and 0.76, respectively) or phenanthrene (KIE = 0.96 and 1.02, respectively) (Table 2).

Mechanistic Significance of Inverse Isotope Effects

The observation of a significant inverse isotope effect in the anthracene oxidation by FePcS/H2O2 or FeTMPS/ KHSO₅ indicated that these KIEs could not be interpreted as primary KIEs involving a C-H(D) bond cleavage in the rate determining step of the initial step of the oxidation reaction. Consequently, these inverse KIEs precluded an oxygen rebound mechanism which involves a hydrogen atom abstraction from substrate by an electrophilic highvalent oxoiron complex to give a short-life radical intermediate followed by rapid recombination step^[23]. Inverse KIEs are usually associated with secondary KIEs when no C-H(D) bond is cleaved or formed, but this bond should modify some parameters in the transition state (TS). This is usually the case when a C-H(D) bond is located at the reaction site itself, containing the leaving group (α secondary KIE) or at the adjacent carbon atom (β secondary KIE). In the case of the intermolecular [H₁₀]anthracene/ $[D_{10}]$ anthracene competitive oxidation an α secondary KIE is observed. Generally, the magnitude of an α -secondary KIE is determined by an inverse stretching and a normal bending contribution to the KIE value [24][25]. For example, in S_N2 reactions, a tighter TS, with short nucleophile α carbon and/or α-carbon leaving group bond lengths, the KIE values would be inverse or normal and small^[24]. Based on calculations with model systems, the α -secondary KIEs have been interpreted as reflecting the changes in the outof-plane bending motion, i.e. hybridization changes or differences in the loose/tight character of the TS^[26]. The activation energy for $sp^2 \rightarrow sp^3$ change in hybridization is lower for the deuterated substrate compared to the non-deuterated analogue, creating then an inverse KIE^[27]. When the TS of the reaction occurs very late along the reaction coordinate, i.e. the TS exhibits essentially a fully sp³-hybridized

carbon atom like in the reaction product, the $k_{\rm H}/k_{\rm D}$ value will be more pronounced, the maximum KIE expected for a sp² \rightarrow sp³ rehybridization will be 0.71^[27]. Values of $k_{\rm H}/$ $k_{\rm D}$ ranging from 0.8 to 0.9 correspond to TS geometries somewhere between sp² and sp³ and are more frequently observed. Inverse secondary KIEs have been measured for the addition of a variety of electrophilic reagents to olefins and aromatics, e.g. for the reaction of 1-naphthylcarbene to toluene $(k_H/k_D = 0.52)^{[28]}$. An other example of an inverse α -secondary $k_{\rm H}/k_{\rm D}$ equal to 0.83 has been recently observed during of the [2 + 2] photocycloaddition of arylalkenes to C₆₀^[29]. Inverse KIEs have been reported for styrene epoxidations catalyzed by cytochrome P-450 ($k_{\rm H}/k_{\rm D}=0.93$) or performed with m-chloroperbenzoic acid (k_H/k_D) $(0.82)^{[27a]}$ or by hexacyanoferrate(III) in acid medium (k_H / $k_{\rm D} = 0.80)^{[27b]}$. The cytochrome P-450 catalyzed oxidation of isotopically labeled chlorobenzenes provides also an inverse KIE (0.95)[30]. This result has been explained by an initial attack of an activated triplet-like oxygen atom on the π -system of substrate to generate a tetrahedral intermediate followed by rearrangement to phenol directly or via epoxide or ketone intermediates^[30]. The aromatic hydroxylation of o- and p-xylene, catalyzed by cytochrome P-450, showed also an inverse α -secondary isotope effects (0.83–0.94) which were consistent with an addition reaction of a oxoiron species onto an sp²-hybridized carbon atom^[31]. The inverse nature of KIEs was anticipated for sp²-to-sp³ rehybridization during the covalent modification of human 5α -reductases by finasteride (a specific inhibitor) by a nucleophilic N addition to the double bond of this steroid-type molecule^[32]. The reaction of the nitroethane anion (the α carbon atom is sp²-hybridized) with D-amino acid oxidase proceeds with an inverse $k_{\rm H}/k_{\rm D}=0.84$ which was indicative of significant sp²-to-sp³ rehybridization in the TS^[33]. Apart from the sp²-to-sp³ rehybridization, inverse isotope effects can be due to the difference in C-H and C-D distances, C-D bond being slightly shorter than corresponding C-H bond (d = 1.112 and 1.107 Å, respectively)^[34]. The racemization of 2,2'-dibromo-4,4'-dicarboxybiphenyl[35] and 9,10dihydro-4,5-dimethylphenanthrene^[36] showed inverse KIEs of 0.85 and 0.88, respectively. The origin of a large inverse secondary KIE in the bromination of a sterically congested olefin (0.36-0.65) has been attributed to the fact that C-D bonds are shorter and less sterically demanding^[37]. Large inverse equilibrium isotope effects have also been observed for the oxidative addition of H_2 to trans-W(PMe₃)₄ I_2 (K^H_{eq}) $K^{\rm D}_{\rm eq}=0.63)^{[38a]}$ and cyclohexane to CpRh(CO)₂ ($K^{\rm H}_{\rm eq}/K^{\rm D}_{\rm eq}=0.1$, at 163–193 K)^[38b]. Recently, a substantial inverse isotope effect of $k_{\rm H}/k_{\rm D}=0.48$ has also been published in Co^{III}-catalyzed polymerization of ethylene^[39] and explained by the presence of a β-agostic Co···H···C (Co···D···C) interaction, the inverse KIE being associated with the release of the agostic bond. Related to this interpretation of an inverse KIE, it must be noted that complexation of n-heptane with double A-frame porphyriniron(III) has been recently observed in crystals^[40]. An out-of-plane displacement of the iron center of 0.26 Å and an Fe···C distance of 2.5 Å strongly suggest moderate to

weak agostic interactions between C-H bonds of the alkane and the metal center. *p*-Xylene can interact with the metal center of an porphyriniron(III), the smallest Fe-C distance was 2.94 Å and the iron atom was drawn 0.07 Å out of the mean plane of the porphyrin core indicating a possible weak agostic interaction^[41].

In the present case of the oxidation of anthracene catalyzed by phthalocyanine- or porphyriniron complexes, anthracene, a polyaromatic substrate, should have a pronounced tendency to form a stacking complex with FePcS or FeTMPS. A rather polar reaction medium (an acetonitrile/water mixture) should favor stacking interactions between two large planar hydrophobic molecules. Then a question arises: can the difference in C-H/C-D substitution on anthracene generate different stacking interactions between [H₁₀]anthracene or [D₁₀]anthracene with the macrocyclic plane molecule of FePcS or FeTMPS? In other words, are the observed KIEs only related to hydrophobic interactions between the aromatic substrate and the macrocyclic catalyst? The differences observed on inverse KIEs during the [H₁₀]anthracene/[D₁₀]anthracene and [H₁₀]anthracene/[D₂]anthracene competitive oxidations (Table 2) could be due to possible differences in stacking interactions between [D₁₀]anthracene and [D₂]anthracene with the macrocycle of the catalyst resulting from a slight difference of the nature of C-H bonds compared to C-D bonds. So, the stacking interactions should increased with the deuterium content of anthracene. This is the case with both catalytic systems: the KIE is 0.90 for [D₂]anthracene and 0.75 with [D₁₀]anthracene with FePcS/H₂O₂ (0.95 and 0.76 with FeTMPS/KHSO₅, respectively) (Table 2). Consequently, the value of the inverse KIE due to the sp²-to-sp³ hybridization change should be enhanced in a highly stacked substrate-catalyst intermediate during the addition of an electrophilic oxoiron complex to the sp²-carbon atom of anthracene to give a σ adduct. The data on ¹⁸O labeling experiments are also consistent with this proposal (see below).

The quasi-absence of KIE observed in the phenanthrene oxidation by FePcS/H₂O₂ or by FeTMPS/KHSO₅ (0.96 and 1.02, respectively) can be due to the reduction of stacking interactions compared to anthracene and also to the different nature of C9–H and C10–H bonds in phenanthrene. The C9–C10 bond of phenanthrene exhibits a relatively high bond order, indicating that it possesses a considerable degree of double bond character (K region bonds of polyaromatics), while anthracene bonds show essentially an aromatic character (*meso* region bonds)^[42]. As a result, the main product of phenanthrene oxidation was biphenyl-2,2′-dicarboxylic acid. A similar cleavage of the double bond of styrene was observed when oxidized by FePcS/H₂O₂^[16].

Mechanistic Significance of ¹⁸O Labeling Experiments

In aqueous media oxo(porphyrin)metal(V) complexes have an axial hydroxo ligand. In order to explain the observed 50% ¹⁸O incorporation from H_2 ¹⁸O in epoxides during olefin epoxidations by a metalloporphyrin/KHSO₅ system^{[20a][21b][21c]}, hydroxylation of DNA deoxyriboses^[20b] or

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C-H bond hydroxylation of 4-isopropylbenzoic acid^[20c] it was proposed that the active hydroxo(oxo)(porphyrin)metal(V) complex undergoes a "redox tautomerism" (Figure 4). The ¹⁸O incorporation depends on the reaction conditions^[21b], primarily on relative kinetics of oxidation step and ligand exchange processes between the hydroxo ligand and bulk water and the nature of the catalyst. The rate of the oxo-hydroxo interchange in the oxo(porphyrin)manganese(V) complex was estimated to be about 10³ s⁻¹[^{21c]}.

Figure 4. Redox tautomerism in a hydroxo(oxo)metal(V) species having a water-soluble porphyrin ligand (O = 16 O, black O = 18 O)

$$\begin{array}{c}
 & \text{oxidant-}^{16}O \\
 & \text{M}^{\text{U}}
\end{array}$$

The large ¹⁸O incorporation into anthrone (63%) and anthraquinone (55%) observed during FeTMPS/KHS¹⁶O₅ anthracene oxidation in the presence of H₂¹⁸O is consistent with the involvement of oxo(porphyrin)iron complex as active species. By contrast, the absence of ¹⁸O incorporation into anthrone, produced in the oxidation by FePcS/H₂O₂, could be explained by the involvement of an active oxometal species unable to undergo such an oxo-hydroxo equilibrium (in particular if there is no hydroxo ligand in the axial position *trans* to the oxo group).

Tautomeric Equilibria in Oxidation Products of Anthracene

The sequence of anthracene oxidation to anthraquinone and the relation between tautomeric forms of products are depicted in Scheme 5. The first oxidation product, 9-hydroxyanthracene is in fast keto-enol equilibrium with anthrone, which is the predominant form. The anthrone/9-hydroxyanthracene ratio determined by NMR in CDCl₃ at 25 °C was estimated to be $80:20^{[43]}$. 9,10-Dihydroxyanthracene exists in solution in equilibrium with oxanthrone ($K_{\rm eq}=0.13$) and is the major tautomer (89%)^[44]. However, the equilibrium is reached very slowly. The rate constants for the 9,10-dihydroxyanthracene-oxanthrone interconversion were determined to be $k_{-2}=3\times 10^{-6}~{\rm s}^{-1}$ and $k_2=4\times 10^{-7}~{\rm s}^{-1}$ [45]. It means that in 1 mM solution only 1% of oxanthrone will be transformed into 9,10-dihydroxyanthracene after 1 h at room temperature.

Mechanism of Anthracene Oxidation by FeTMPS/KHSO₅

The oxidation of anthracene by FeTMPS/KHSO₅ was significantly faster than those of anthrone and oxanthrone. Under the same conditions the conversions of anthracene, anthrone and oxanthrone after 30 min were 100, 38 and 18%, respectively. These data indicated that there was an oxidation pathway to give directly oxanthrone, the main oxidation product, without intermediate formation of anthrone.

An electrophilic attack of a high-valent oxo(porphyrin)iron complex at a central sp²-carbon atom of anthracene to

Scheme 5. Oxidation products of anthracene and tautomeric equilibria for anthrone and oxanthrone

give a σ complex is probably the initial oxidation step (Figure 5). The inverse isotope effect, indicative of an sp²-tosp³ rehybridization in the TS containing both substrate and catalyst stacked and the observed ¹⁸O incorporation into anthrone, the first oxidation product, typical for oxoiron complex mediated oxidation in aqueous medium strongly suggested the formation of a new carbon-oxygen bond to form a σ complex A. This intermediate A can be trapped by O₂ to produce oxanthrone and anthraquinone via an intermediate radical B. An intramolecular electron transfer in intermediate A can lead to an intermediate carbocation C, which can undergo either a proton elimination to give a σ complex **D** or a nucleophilic attack of water to produce an intermediate E, providing a reaction pathway for oxanthrone formation without intermediate formation of anthrone. The hydrolysis of the Fe^{III}-O-aromatic bond in both intermediates D and E gave 9-hydroxyanthracene (quickly tautomerized to anthrone) and 9,10-dihydro-9,10dihydroxyanthracene F. Then F can be further oxidized by a 2e⁻ process with the release of 2H⁺, producing oxanthrone with an increased ¹⁸O content, or by oxidation of a C-H bond by an oxygen rebound mechanism. The GC-MS analysis of [H₁₀]anthracene/[D₁₀]anthracene oxidation products after 6 min of reaction revealed that anthraquinone was enriched with nondeuterated material: 57% of [H₈]anthraquinone and 43% of [D₈]anthraquinone, indicating a KIE value > 1 on the anthrone-to-anthraquinone oxidation step. The ¹⁸O content in anthraquinone (55%) was not increased compared to that in anthrone (63%). Both findings suggest that oxidation of intermediate F proceeds by an oxygen rebound mechanism followed by H₂O elimination to generate oxanthrone. The fact that the metallopor-

Figure 5. Proposed mechanism for anthracene oxidation by FeTMPS/KHSO₅ (O = 16 O, black \bullet = 18 O)

phyrin-catalyzed oxidation of anthracene gave oxanthrone, not the more oxidizable 9,10-hydroxyanthracene tautomeric form, explains the rather low yield of anthraquinone, since oxanthrone is relatively stable under the reaction conditions (oxanthrone conversion being only 23% after 60 min).

Active Species in FePcS/H2O2 Oxidations

In the FePcS/H₂O₂ mediated oxidation of trichlorophenol^[15][17] a nucleophilic PcSFe^{III}OO⁻ complex was proposed to be the active species, able to perform two key steps: (i) a nucleophilic addition to the electron-deficient double bond of intermediate quinones to provide epoxy quinones and (ii) a nucleophilic attack on the carbonyl group of the different quinones, leading to their ring cleavage^[16]. Although the addition of nucleophiles to polyar-

enes, e.g. the addition of alkyllithium reagents to anthracene, has been published^[46], such a peroxoiron(III) complex is probably not nucleophilic enough for a direct attack onto an sp²-carbon atom of a PAH. Another possible active metal-oxygen species could be a high-valent oxoiron complex, having two redox equivalents above the Fe^{III} state. Several lines of evidence indicate that it is not the case for the FePcS/H₂O₂ system^[15b]. The fact that the oxygen atom in anthrone originated from H₂O₂ (Figure 3) also indicated that an oxoiron(V) complex was not involved in anthracene oxidation by FePcS/H₂O₂. With such an oxoiron(V) complex we could expect both ¹⁶O and ¹⁸O incorporations due to redox tautomerism, as observed in anthracene oxidation by FeTMPS/KHSO₅. The other possible formulation of the active species involved in PAH oxidations by FePcS/H2O2 is a PcSFe^{IV}=O species^[15b]. It can be formed by a 1e⁻ oxi-

Figure 6. Generation of peroxoiron(III) and oxoiron(IV) species by activation of FePcS with hydrogen peroxide

dation of PcSFe^{III}OH complex (Figure 6, pathway A) or by the intermediate formation of a μ -peroxo dimer (pathway B). In the former case ¹⁸O from water should be found in anthrone obtained in the presence of H2¹⁸O. The absence of ¹⁸O incorporation into anthrone during the oxidation of anthracene by FePcS/H₂O₂ in H₂¹⁸O eliminated this possibility. Consequently, PcSFe^{IV}=O should be formed by pathway B by the homolytic cleavage of a μ -peroxo dimer (we proposed the pathway A in ref.^[15b] before having on hands the data of these labeling studies). This PcSFe^{IV}=O species is neutral and does not require the presence of an anionic axial anion ligand (e.g. HO⁻) to compensate a charge. Consequently, no redox tautomerism is possible in the absence of HO⁻ ligand *trans* to the oxo entity. This

absence of redox tautomerism for $PcSFe^{IV} = O$ explained the 100% incorporation of ^{16}O into anthrone during the FePcS/ H_2O_2 oxidation of anthracene performed in heavy water.

Mechanism of Anthracene Oxidation by FePcS/H2O2

The first step of the FePcS/H₂O₂ mediated oxidation of anthracene resembles that described in the FeTMPS/ KHSO₅ oxidation (Figure 7). The nearly identical inverse KIE (0.75) found for the first reaction step suggested also the formation of a σ complex, intermediate A' which can be trapped by O2 to produce oxanthrone and anthraquinone via a radical intermediate B'. The significant ¹⁶O incorporation in anthraquinone when the oxidation was performed under air compared to that obtained under nitrogen (Figure 3) was strongly indicative of this pathway 1. Hydrolysis of the Fe^{III}-O-aromatic bond followed by proton elimination and 1e⁻ oxidation generated 9-hydroxyanthracene which rapidly tautomerized to the thermodynamically favored anthrone. Successive 1e⁻ oxidation and proton elimination steps gives rise to carbocation I', which can react with water (pathway 2) to produce oxanthrone and anthraquinone. PcSFe^{IV}=O complex is not such a strong oxidant to perform the abstraction of an H atom of a aliphatic or aromatic C-H bond. However, PcSFe^{IV}=O is

Figure 7. Proposed mechanism for anthracene oxidation by FePcS/H₂O₂ (O = 16 O, black \bullet = 18 O)

probably a sufficiently good 1e⁻ oxidant to perform 1e⁻ oxidations of intermediate products or of oxanthrone by successive 1e⁻ oxidations and proton eliminations to provide anthraquinone.

The rather low yields of anthrone and anthraquinone after 30 min (5 and 21%, respectively) might be due to radical intermediates capable of undergoing side reactions, e.g. coupling reactions. Accordingly, the presently proposed mechanism, the first oxygen atom incorporated into the different products originates from PcSFe^{IV}=O (¹⁶O in labeling experiments), and the value of ¹⁸O incorporation of the second oxygen atom of anthraquinone from H₂¹⁸O will depend on the relative rates of the different pathways indicated on Figure 7. Very significant ¹⁸O incorporation into the second oxygen position of anthraquinone under air (64%) and, especially under nitrogen (88%), strongly suggest that pathway 2 is the major reaction pathway.

Conclusion

The (tetrasulfophthalocyanine)iron/hydrogen peroxide oxidation system, based on an easy-to-prepare catalyst and a "clean" oxidant, is capable of oxidizing polycyclic aromatic hydrocarbons, like anthracene, phenanthrene and benzo[a]pyrene. Data obtained from product distribution, kinetic isotope effects and ¹⁸O labeling studies, as well as a comparative study of anthracene oxidation by FeTMPS/ KHSO₅ allowed us to propose that an oxo(phthalocyanine)iron(IV) complex, is probably responsible of these PAH oxidations. Consequently, taking in consideration the previously reported mechanistic studies on quinone cleavage by the same FePcS/H₂O₂ system, two kinds of the active species can be generated from FePcS in the presence of H₂O₂: (i) a nucleophilic peroxoiron complex PcSFe^{III}OO⁻ responsible for double carbon-carbon bond cleavage in the deep oxidation of trichlorophenol^{[15][16]} and (ii) an electrophilic PcSFe^{IV}=O complex capable of oxidizing aromatic hydrocarbons by the formation of a σ adduct. This versatility makes (tetrasulfophthalocyanine)iron in association with hydrogen peroxide a suitable catalytic method for the chemical degradation of different pollutants.

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Experimental Section

Instrumentation: Gas chromatography-mass spectrometry (GC-MS) was performed with a Hewlett-Packard 5890 instrument using electron-impact ionization at 70 eV. The carrier gas for GC-MS was He, and a 12 m \times 0.2 mm HL-1 (Crosslinked Methyl Silicone Gum) capillary column was used. — High performance liquid chromatography (HPLC) analyses were performed with a Waters-486 Liquid Chromatograph equipped with a μ -Bondapak C18 column, eluent being a mixture of methanol and water (85:15, v/v) at a flow rate of 1 ml min $^{-1}$, detection at 242 nm for anthracene, 252 nm for phenanthrene and 280 nm for benzo[a]pyrene analyses. — UV/Vis spectra were recorded with a Hewlett-Packard 8452A diode array spectrophotometer. — 1 H NMR spectra were recorded with Bruker WM 250 spectrometer.

Materials: All chemicals used were of reagent grade. Potassium monopersulfate was a gift from Interox (Curox^R) and is the triple salt 2KHSO₅·KHSO₄·K₂SO₄. Hydrogen peroxide was obtained from Janssen Chimica as a 35% aqueous solution. H₂¹⁸O (98 atom-%) was supplied by Eurisotop (Gif-sur-Yvette, France). Lithium aluminum deuteride was purchased from Fluka (minimum 99 atom-% D). (Tetrasulfophthalocyanine)iron (FePcS) was prepared according to the method of Weber and Busch. [47] The iron complex octasodium *meso*-tetrakis(3,5-disulfonatomesityl)porphyrin (TMPS) was synthesized according to ref.^[48]. Anthracene (97%), phenanthrene (98%), benzo[a]pyrene (98%), [D₁₀]anthracene (> 98 atom-% D), [D₁₀]phenanthrene (97 atom-% D), anthrone (96%), anthraquinone (97%), phenanthrenequinone (99%), 9,10-dibromoanthracene (98%) were purchased from Aldrich. Oxanthrone was prepared from anthrone via 10-bromoanthrone according to ref. [45]. NMR (CD₂Cl₂): $\delta = 2.47$ (d, 1 H, J = 9.5 Hz, OH), 5.78 (d, 1 H, $J = 9.5 \text{ Hz}, H_{10}$, 7.54 (td, 2 H, J = 7.5 and 0.7 Hz, H₃), 7.73 (td, 2 H, J = 7.5 and 1.5 Hz, H₂), 7.90 (dd, 2 H, J = 7.5 and 0.7 Hz, H_4), 8.25 (dd, 2 H, J = 7.5 and 1.5 Hz, H_1); (CD₂Cl₂ + small amount of D_2O): the same signals, except doublet at $\delta = 2.47$ disappeared and the doublet at $\delta = 5.78$ became a singlet.

Preparation of 9,10-Dideuterioanthracene: The procedure was adapted from ref.^[49]. Dibromoanthracene (1.008 g, 3 mmol) and Li-AlD₄ (504 mg, 12 mmol) were added under nitrogen to 15 ml of anhydrous dimethoxyethane. The reaction mixture, contained in a 50-ml round-bottom single-neck flask and maintained under nitrogen, was partly submerged in a ultrasonic laboratory cleaner Branson 2200 (working frequency 47 kHz, 205 W) at 35-40°C. The reaction was monitored by GC-MS. After 13 h, all dibromoanthracene was reduced and the reaction mixture contained 9,10-dideuterioanthracene accompanied by minor amount of 9-bromo[D₁]anthracene. The reaction mixture was cooled at -40°C and 1 ml of D₂O (99%) was slowly added. The temperature was allowed to return to 20°C and hydrolysis of the excess of LiAlD₄ was performed for 30 min. Then the resulting mixture was pourred into 30 ml of water and extracted with diethyl ether (4 × 40 ml). The combined organic extracts were washed with saturated NaCl solution, dried with Na₂SO₄ and concentrated. The white precipitate was separated by filtration and recrystallized from diethyl ether to give 148 mg (27% yield) of the pure 9,10-dideuterioanthracene material. The isotopic purity was determined by NMR and GC-MS methods. The material contained 72.8% of 9,10-dideuterioanthracene and 27.2% of 9-deuterioanthracene.

General Catalytic Procedure for PAH Oxidation: Oxidation of anthracene was carried out in a mixture of acetonitrile/50 mm phosphate buffer at pH = 7 (3:1, v/v) at 20° C under aerobic conditions. Oxidations of phenanthrene and benzo[a]pyrene were performed in acetonitrile/33.3 mm phosphate buffer at pH = 4.6 (1:1, v/v). The reaction mixture (2 ml) contained 2 µmol of substrate introduced as a 4 mm acetonitrile solution and 0.074 or 0.1 µmol of catalyst, introduced as a 1.28 mm solution in water, and the required amounts of acetonitrile and stock buffer solution to obtaine the desired organic solvent/water ratios. Oxidations were started by addition of 10 µmol of oxidant (10 µl of 3.5% H₂O₂ solution or 3.1 mg of potassium monopersulfate). The final concentrations of the catalyst, substrate and oxidant were 37 (or 100) µm, 1 mm and 5 mm, respectively. The reactions were followed by HPLC and GC-MS techniques. Anthraquinone and anthrone were identified by comparison of its GC-MS behavior (retention time and mass spectrum) with those of an authentic sample. Oxanthrone was identified by comparison of its NMR spectrum with that of an authentic sample.

 $H_2^{18}O$ Experiments: Reactions were carried out using $H_2^{18}O$ (98) atom-%) according to the following procedure. A mixture of 25 µl of 0.5 M phosphate buffer pH = 7 and 29 μ l of catalyst stock solution (1.28 mm in water) was dried under vacuum. The dry residue was then dissolved in 250 µl of H₂¹⁸O and 250 µl of [H₁₀]anthracene and [D₁₀]anthracene stock solutions (50:50 mixture, 4 mm total concentration in MeCN) and 500 µl of MeCN were added. The oxidations were performed under air and under nitrogen. In the latter case the reaction mixture was submitted to 3 freeze-pump-thaw cycles and the reaction flask was filled up with nitrogen. The reaction was started by the addition of 5 µl of 3.5% H₂O₂ (for FePcS oxidation) or 1.6 mg of KHSO₅ (for FeTMPS oxidation). The substrate conversion was monitored by HPLC and isotopic compositions of products were analyzed by GC-MS.

Determination of Isotopic Composition of Substrates and Products: Mass spectra of [H₁₀]anthracene and [D₁₀]anthracene show very strong molecular peaks accompanied with little fragmentations. [H₁₀]anthracene: MS; m/z (%): 178 (100) [M⁺], 176 (18) [M $-2 H]^+$, 152 (8) [M $-C_2H_2]^+$, 89 (13) [M $-C_7H_5]^+$. [D₁₀]anthracene: MS; m/z (%): 188 (100) [M⁺], 184 (13) [M - 2 D]⁺, 160 (9) $[M - C_2D_2]^+$, 94 (17) $[M - C_7D_5]^+$. So, under conditions of GC-MS analysis, within experimental error, there is no isotope effect on fragmentation and isotopic compositions of [H₁₀]anthracene and [D₁₀]anthracene can be determined from the intensities of corresponding molecular peaks. The mass spectra of phenanthrene and products of oxidation of anthracene and phenanthrene showed the same features and their isotopic compositions were determined from the intensities of the molecular peaks of deuterated and nondeuterated compounds.

KIE determinations based on [H₁₀]anthracene/9,10-dideuterioanthracene oxidations were performed taking into account admixture of monodeuterated anthracene, the presence of [M + 1] peaks due to 13 C natural content and [M - 1] and [M - 2] fragmentations according to the following formalism:

$$I_{178} = [D_0]-A + F_{M-1}\cdot[D_1]-A + F_{M-2}\cdot[D_2]-A$$
 (2)

$$I_{179} = [D_1]-A + F_{M+1}\cdot[D_0]-A + F_{M-1}\cdot[D_2]-A$$
 (3)

$$I_{180} = [D_2] - A + F_{M+1} \cdot [D_1] - A$$
 (4)

where I_{178} , I_{179} , I_{180} were intensities of peaks at m/z 178, 179 and 180, respectively; [D₀]-A, [D₁]-A, [D₂]-A were percentages of corresponding isotopomers of anthracene; F_{M+1} , F_{M-1} , F_{M-2} were relative intensities of [M + 1], [M - 1] and [M - 2] peaks. The latter intensities were determined in separate GC-MS experiments to be $F_{M+1} = 0.15$, $F_{M-1} = 0.11$, $F_{M-2} = 0.18$. KIEs were calculated from the data obtained on conversions and the percentages of [H₁₀]anthracene and 9,10-dideuterioanthracene in an initial reaction mixture and after several minutes when the substrate conversion was near 50%.

Biol. Chem. 1986, 261, 6900-6903.

K. E. Hammel, B. Kalyanaraman, T. K. Kirk, J. Biol. Chem. **1986**, *261*, 16948–16952

[5] J. D. Lipscomb, A. M. Orville in Degradation of Environmental Pollutants by Microorganisms and their Metalloenzymes. Metal Ions in Biological Systems (Eds.: H. Sigel, A. Sigel), Marcel Dekker, New York, 1992, vol. 28, p. 243.

D. A. Lane in *Chemical Analysis of Polycyclic Aromatic Compounds* (Ed.: T. Vo-Dinh), Wiley, New York, 1989, p. 31.
 For a review on chemistry of polyarenes see: R. G. Harvey, *Polycyclic Aromatic Compounds*, Wiley, New York, 1997, p. 96.

M. Juaristi, J. M. Aizpurua, B. Lecea, C. Palomo, Can. J. Chem. **1984**, *62*, 2941-2944

[9] E. V. Dehmlov, J. K. Makrandi, J. Chem. Research (S) 1986, 32 - 33.

[10] D. Villemin, M. Ricard, Nouv. J. Chem. 1984, 8, 185-189.

[11] W. Adam, P. A.Ganeshpure, *Synthesis* **1993**, 280–282.

[12] [12a] R. Song, A. Sorokin, J. Bernadou, B. Meunier, *J. Org. Chem.* **1997**, 62, 673–678. – [12b] G. J. Harden, M. M. Coombs, J. Chem. Soc., Perkin Trans. 1 1995, 3037–3042.

[13] T. Higuchi, C. Satake, M. Hirobe, J. Am. Chem. Soc. 1995, 117, 8879-8880.

[14] S. V. Barkanova, V. M. Derkacheva, O. V. Dolotova, V. D. Li, V. M. Negrimovsky, O. L. Kaliya, E. A. Luk'yanets, *Tetrahedron Lett.* **1996**, *37*, 1637–1640.

[15] [15a] A. Sorokin, J.-L. Séris, B. Meunier, Science 1995, 268, 1163–1166. – [15b] B. Meunier, A. Sorokin, Acc. Chem. Soc. 1997, 30, 470–476.

 [16] [16a] A. Sorokin, B. Meunier, *Chem. Eur. J.* 1996, 2, 1308-1317.
 - [16b] A. Sorokin, S. De Suzzoni-Dezard, D. Poullain, J.-P. Noël, B. Meunier, *J. Am. Chem. Soc.* 1996, 118, 7410-7411.
 - [16c] A. Hadasch, A. Sorokin, A. Rabion, B. Meunier, *New J.* Chem. 1998, 45-51.

[17] A. Sorokin, L. Fraisse, A. Rabion, B. Meunier, J. Mol. Cat. **1997**, 117, 103-114.

[18] R. J. Lorentzen, J. Caspary, S. A. Lesko, P. O. Ts'o, *Biochemistry* **1975**, *14*, 3970–3977.

L. Melander, J. M. Saunders, Reaction Rates of Isotopic Molecules, Wiley, New York, 1980, p. 95.

ecules, Wiley, New Tork, 1700, p. 23.

[20] [20a] J. Bernadou, A.-S. Fabiano, A. Robert, B. Meunier, J. Am. Chem. Soc. 1994, 116, 9375–9376. – [20b] M. Pitié, J. Bernadou, B. Meunier, J. Am. Chem. Soc. 1995, 117, 2935–2936. – [20c] R. J. Balahura, A. Sorokin, J. Bernadou, B. Meunier, Inorg. Chem. 1997, 36, 3488-3492.

[21] [21a] W. Nam, J. S. Valentine, *J. Am. Chem. Soc.* **1993**, *115*, 1772–1778. – [21b] K. A. Lee, W. Nam, *J. Am. Chem. Soc.* **1997**, *119*, 1916–1922. – [21c] J. T. Groves, J. Lee, S. S. Marla, J. Am. Chem. Soc. 1997, 119, 6269-6273.

[22] B. Meunier, Chem. Rev. 1992, 92, 1411-1456.

 [23] [23a] T. J. McMurry, J. T. Groves, In Cytochrome P-450 Structure, Mechanism and Biochemistry (Ed.: P. R. Ortiz de Montellano), Plenum, New York, 1986; pp. 1-28. — [23b] M. Newcomb, M. V. J. T. V. Biodzie, D. L. Chastray, E. S. Boberte, P. E. Holling, P. L. Chastray, E. S. Boberte, P. E. Holling, P. L. Chastray, E. S. Boberte, P. E. Holling, P. L. Chastray, E. S. Boberte, P. E. Holling, P. L. Chastray, E. S. Boberte, P. E. Holling, P. L. Chastray, E. S. Boberte, P. E. Holling, P. L. Chastray, E. S. Boberte, P. E. Holling, P. L. Chastray, E. S. Boberte, P. E. Holling, P. L. Chastray, E. S. Boberte, P. E. Holling, P. L. Chastray, E. S. Boberte, P. E. Holling, P. L. Chastray, E. S. Boberte, P. E. Holling, P. L. Chastray, P. E. Holling, P. L. Chastray, P. E. Holling, P. L. Chastray, P. E. Chastray, P. E. Chastray, P. E. L. Chastray, P. E. Chastray, P. Chastray, P. E. Chastray, P. E. Chastray, P. E. Chastray, P. E. C H. Le Tadic-Biadatti, D. L. Chestney, E. S. Roberts, P. F. Hollenberg, J. Am. Chem. Soc. 1995, 117, 12085–12091.

[24] R. A. Poirier, Y. Wang, K. C. Westaway, J. Am. Chem. Soc. 1994,

116, 2526-2533

[25] S. Wolfe, C.-K. Kim, J. Am. Chem. Soc. **1991**, 113, 8056–8061. [26] [26a] P. A. Nielsen; S. S. Glad, F. Jensen, J. Am. Chem. Soc. 1996,
 118, 10577-10583. - [26b] S. S. Glad, F. Jensen, J. Am. Chem.

Soc. 1997, 119, 227–232.

[27] [27a] R. P. Hanzlik, G. O. Shearer, Biochem. Pharm. 1978, 27, 1441–1444 and references therein. – [27b] A. K. Bhattacharjee, M. K. Mahanti, Gazz. Chim. Ital. 1984, 114, 337-340

[28] G. W. Griffin, K. A. Horn, J. Am. Chem. Soc. 1987, 109, 4919-4926 and references therein.

[29] G. Vassilikogiannakis, M. Orfanopoulos, J. Am. Chem. Soc. **1997**, *119*, 7394–7395.

[30] K. R. Korzekwa, D. C. Swinney, W. F. Trager, Biochemistry 1989, 28, 9019-9027.

[31] R. P. Hanzlik, K.-H. J. Ling, J. Am. Chem. Soc. 1993, 115, 9363 - 9370.

[32] G. Tian, S.-Y. Chen, K. L. Facchine, S. R. Prakash, J. Am. Chem. Soc. 1995, 117, 2369-2370.

K. A. Kurtz, P. F. Fitzpatrick, J. Am. Chem. Soc. 1997, 119, 1155 - 1156.

[34] J. March, Advanced Organic Chemistry: Reactions, Mechanisms and Structure, 4th ed., Wiley, New York, 1992, p. 20.

[35] L. Melander, R. E. Carter, Acta Chem. Scand. 1964, 18, 1138-1149.

[36] K. Mislow, R. Graeve, A. J. Gordon, G. H. Wahl, Jr., J. Am.

Chem. Soc. 1963, 85, 1199–1200.

R. W. Nagorski, H. Slebocka-Tilk, R. S. Brown, J. Am. Chem. Soc. 1994, 116, 419–420.

[38] [38a] D. Rabinovich, G. Parkin, J. Am. Chem. Soc. 1993, 115, 353-354. - [38b] R. H. Schultz, A. A. Bengali, M. J. Tauber, B. H. Weiller, E. P. Wasserman, K. R. Kyle, C. B. Moore, R. G. Bergman, *J. Am. Chem. Soc.* **1994**, *116*, 7369–7377.

[39] M. J. Tanner, M. Brookhart, J. M. DeSimone, J. Am. Chem. Soc. 1997, 119, 7617-7618.

^[1] C. E. Cerniglia, *Biodegradation* **1992**, 3, 351–368.
^[2] K. E. Hammel in *Degradation of Environmental Pollutants by* Microorganisms and their Metalloenzymes. Metal Ions in Biological Systems (Eds.: H. Sigel, A. Sigel), Marcel Dekker, New York, **1992**, vol. 28, p. 41. S. D. Haemmerli, M. S. A. Leisola, D. Sanglard, A. Fiechter, *J.*

- [40] D. R. Evans, T. Drovetskaya, R. Bau, C. A. Reed, P. D. W. Boyd, J. Am. Chem. Soc. 1997, 119, 3633-3634.
 [41] Z. Xie, R. Bau, C. A. Reed, Angew. Chem. Int. Ed. Engl. 1994, 33, 2433-2434.
 [42] See ref. [7], p. 26.
 [43] T. R. Criswell, B. H. Klanderman, J. Org. Chem. 1974, 39, 770-774.
 [44] K. Brederesk, E. F. Sommermann, Tetrahedron Lett. 1966, 41, 5009-5014

- [45] S. A. Carlson, D. H. Hercules, *Anal. Chem.* **1973**, 45, 1794–1799.

- [794-1799.]
 [46] P. P. Fu, R. G. Harvey, Chem. Rev. 1978, 78, 317-361.
 [47] J. H. Weber, D. H. Busch, Inorg. Chem. 1965, 4, 469-471.
 [48] P. Hoffmann, G. Labat, A. Robert, B. Meunier, Tetrahedron Lett. 1990, 31, 1991-1994.
 [49] B. H. Han, P. Boudjouk, Tetrahedron Lett. 1982, 23, 1643-1646.

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